

PII: S0040-4039(96)02408-2

Enantioselective Synthesis of the Prelog-Djerassi Lactonic Acid *via* Group-Selective Aldolization/Desymmetrization of a *Meso* Dialdehyde with a Chiral *N*-Propionylsultam.

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Abstract: The group-selective aldolization/desymmetrization of *meso* dialdehyde 5 with a borylenolate derived from *N*-propionylbornanesultam *ent-2* yields very efficiently lactols 6 with simultaneous generation of four stereogenic centers. Oxidation $(6 \rightarrow 7)$ followed by saponification of the sultam moiety $(7 \rightarrow 4)$ provided the Prelog-Djerassi lactonic acid 4 in a three step sequence in 61-71% overall yield. © 1997, Elsevier Science Ltd. All rights reserved.

The architectural and stereochemical complexity of polyketide-derived natural products offers a unique platform for the development of new synthetic methods and concepts.¹⁾ Among these, new versions of the aldol reaction constitute one of the most powerful ways for generating the contiguous stereogenic centers during the carbon-carbon bond forming steps.²⁾ In this context, we have described the preparation of crystalline, optically pure *syn*- or *anti*-aldols from sultam-derived enolates.³⁾ Furthermore, as part of a recent synthesis of the marine polypropionates (-)-denticulatins A and B, we presented, to the best of our knowledge, for the first time an enantiotopic group differentiation in a *meso* dialdehyde by an aldolization reaction (Scheme 1).^{4,5)} Scheme I

Thus, desymmetrization of *meso* dialdehyde 1 (SiR₃ = TBS or TIPS) by aldolization with the Z-(O)-borylenolate derived from N-propionylbornanesultam 2, furnished a mixture of lactols 3 in 74-95% yield and with diastereomeric ratio's of 11.5-20 : 1. During the process, five contiguous stereogenic centers were generated. Herein we wish to report another application of this approach towards the synthesis of the Prelog-Djerassi lactonic acid 4 (Scheme 2).

Scheme 2

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The Prelog-Djerassi lactonic acid 4 was isolated as an oxidative degradation product of neomethymycin, methymycin, narbomycin and picromycin. (a) It's full stereochemistry was established by Rickards and Smith in 1970. Being the target for numerous synthetic investigations, the Prelog-Djerassi lactonic acid 4, served both in the structural elucidation and the synthesis of macrolide antibiotics. Since substituents and stereochemical interrelationships present in 4 are also found in other oxygenated natural products, the synthesis of Prelog-Djerassi lactonic acid served as a probe for the invention and development of new methods and strategies for controling the stereoselective construction of cyclic and acyclic systems containing three or more contiguous and/or alternating stereocenters. Our very different strategy forsees the desymmetrization of a meso dialdehyde with a chiral sultam-derived enolate (Scheme 2, $5 + II \rightarrow I$) with simultaneous generation of four stereogenic centers. Functional group transformation of aldol I, containing the complete carbon skeleton of 4, would ultimately lead to the Prelog-Djerassi lactonic acid.

In Scheme 3 we outline our synthesis of the Prelog-Djerassi lactonic acid, which began with the crucial aldolization/desymmetrization as a key step.

(a) ent-2 (1 mol-equiv.), Et₂BOTf (1.15 mol-equiv.), i-Pr₂NEt (1.17 mol-equiv.), CH₂Cl₂, 0°C, 0.5 h; 5 (1.5 mol-equiv.), CH₂Cl₂, -78°C, 2 h. (b) TPAP (0.05 mol-equiv.), NMO (1.5 mol-equiv.), powdered 4 Å molec. sieves, CH₂Cl₂, r.t., 2 h. (c) LiOH.H₂O (1.5 mol-equiv.), H₂O₂ (2 mol-equiv.), THF/H₂O (3:1), 0°C, 50 min.

Thus, when N-propionylbornanesultam ent- 2^{3a} (1 mol-equiv.) was successively treated with (freshly prepared and distilled) diethylboryl triflate⁹⁾ (1.15 mol-equiv.)/i-Pr₂NEt (1.17 mol-equiv.) and meso dialdehyde 5^{10} (1.5 mol-equiv.), we obtained after workup and flash chromatography (FC) a mixture of lactols 6 in 75-88% yield. It turned out to be impossible to determine the presence of other stereoisomers at this stage (analysis by ¹H NMR or GC).

We therefore decided to oxidize the mixture of lactols to the corresponding lactones 7 and 8. Oxidation of the mixture of lactols 6 with either $Br_2/NaOAc$, PCC or PDC gave invariable a mixture of lactones 7 and 8, together with tricarbonyl compound 10, obtained *via* oxidation of the open chain alcohol 9 (ratio 7 + 8 : 10 = 1.3-2.6:1, 90% combined yield). However, when the oxidation was carried out with tetrapropylammoniumper-

ruthenate (TPAP)¹¹⁾ in the presence of *N*-methylmorpholine-*N*-oxide and powdered 4 Å molecular sieves in CH₂Cl₂, the α , β -syn- β , γ -anti ("anti-Felkin")^{2b)} product 7 {m.p. 101-102°C, [α]_D = -68.8 (c=1.04, CHCl₃)} was isolated in 75% yield together with the α , β -syn- β , γ -syn ("Felkin") product 8 in 11% yield and no sign of the tricarbonyl compound 10 could be detected. (12) The "anti-Felkin": "Felkin" selectivity (ratio 7: 8) was determined in every case by ¹H NMR analysis of the crude reaction mixture and proved to be invariable to the oxidation conditions (d.r = 7-8:1). We can therefore conclude that the diastereomeric ratio determined for the lactones reflects the stereoselectivity for the aldol reaction/desymmetrization (*ent-2* + 5 \rightarrow 6). Finally, hydrogen peroxide assisted saponification of lactone 7 with lithium hydroxide (1.5 mol-equiv.)/hydrogen peroxide (2 mol-equiv.) in a 3:1 mixture THF/H₂O, furnished the Prelog-Djerassi lactonic acid 4 {m.p. 124-125°C, [α]_D = +39.8 (c=0.71, CHCl₃)} in 94% yield together with a nearly quantitative recovery of the bornanesultam auxiliary.

In summary, the enantioselective synthesis of the Prelog-Djerassi lactonic acid 4, obtained from *meso* dialdehyde 5 via a 3 step sequence in 61-71% overall yield, demonstrates once more the power of the group selective aldolization/desymmetrization with bornanesultam-derived enolates as very efficient chirophores. This represents, to our knowledge, the shortest synthesis of the Prelog-Djerassi lactonic acid. Generalization of this concept with other types of *meso* dialdehydes and differently substituted bornanesultam-derived enolates will be reported in due course.

Financial support of this work by the Swiss National Science Foundation, Sandoz Pharma Ltd, Basel and Givaudan-Roure AG, Dübendorf, is gratefully acknowledged. We thank the Stipendienfonds der Basler Chemischen Industrie for a scholarship to E. W.. We thank Mr. J. P. Saulnier, Mr. A. Pinto and Mrs. D. Klink for NMR and MS measurements.

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- 12) All new compounds were characterized by IR, ¹H-NMR, ¹³C-NMR and mass spectra. Experimental procedure and spectral data for compounds 7 and 8: To a stirred solution of diethylboryl triflate (freshly prepared and distilled⁹, 0.22 ml, 1.22 mmol) in dry CH₂Cl₂ (0.8 ml) was added at 0°C a solution of Npropionyl sultam ent-2 (288 mg, 1.06 mmol) in dry CH₂Cl₂ (1 ml + 1 ml rince) followed by diisopropylethylamine (216 μl, 1.24 mmol). The mixture was stirred for 30 min at 0°C and then cooled to -78°C after which a solution of the meso dialdehyde 5 (204 mg, 1.59 mmol) in CH₂Cl₂ (2 ml + 1 ml rince) was added dropwise at -78°C. After stirring for 1-1.5 h at -78°C, the mixture was quenched by the addition of aq. phosphate buffer (pH 7, 2 ml), allowed to reach r.t., followed by extraction of the aq. phase with CH₂Cl₂ (3 X 10 ml). After drying (MgSO₄) and concentration, the residue was purified by flash chromatography (EtOAc/hexanes 1:4) to yield the corresponding lactols (373 mg, 88%). A Solution of a part of this mixture of lactols (140 mg, 0.35 mmol) in dry CH₂Cl₂ (1 ml) was added to a solution of Nmethyl morpholine-N-oxide (63 mg, 0.54 mmol) and tetrapropylammoniumperruthenate (6.3 mg, 0.017 mmol) in dry CH₂Cl₂ (1 ml), containing 4Å powdered molecular sieves (200 mg). After stirring for 2 h at r.t., the mixture was filtered over a short path of silica gel, concentrated and purified by flash chromatography (silica gel, EtOAc/hexanes 1:5). The major diastereoisomer 7 was isolated in 75% yield (106 mg) and the minor isomer 8 in 11% yield (16 mg): 7: m.p. 101-102°C IR: 2967, 2874, 1725, 1692, 1458, 1376, 1332, 1267 cm⁻¹; ¹H-NMR (400 MHz, CDCl₃) δ 0.98 (s, 3 H), 1.01 (d, 3 H, J = 6.2 Hz), 1.15 (s, 3 H), 1.26 (d, 3 H, J = 6.6 Hz), 1.36 (d, 3 H, J = 7.1 Hz), 1.31-1.46 (m, 3 H), 1.84-2.02 (m, 5 H), 2.02- $2.11 \text{ (m, 2 H)}, 2.45-2.57 \text{ (m, 1 H)}, 3.00-3.35 \text{ (dq, 1 H, } J = 6.6, 7.1 \text{ Hz)}, 3.45 \text{ (d, 1 H, } J = 14.1 \text{ Hz)}, 3.53 \text{ (d, 1 H, } J = 14.1 \text{$ 1 H, J = 14.1 Hz), 3.89 (t, 1 H, J = 6.4 Hz), 4.42 (dd, 1 H, J = 6.6, 8.1 Hz); ¹³C-NMR (50 MHz, CDCl₃) δ 14.6 (q), 17.1 (q), 19.1 (q), 19.9 (q), 20.8 (q), 26.4 (t), 32.7 (d), 32.8 (t), 35.5 (d), 36.9 (t), 38.3 (t), 44.4 (d), 44.5 (d), 47.8 (s), 48.4 (s), 53.2 (t), 65.3 (d), 84.8 (d), 172.8 (s), 174.2 (s). Exact mass calc. for $C_{20}H_{31}O_5NS^+$: 397.1923. Found: 397.1938; **8**: ¹H-NMR (400 MHz, CDCl₃) δ 0.94 (d, 3 H, J = 7.1 Hz), 0.98 (s, 3 H), 1.15 (s, 3 H), 1.19 (d, 3 H, J = 6.6 Hz), 1.44 (d, 3 H, J = 6.6 Hz), 1.00-1.46 (m, 3 H), 1.84-1.001.94 (m, 3 H), 2.00-2.08 (m, 2 H), 2.25-2.47 (m, 2 H), 2.51-2.68 (m, 1 H), 3.38 (dq, 1 H, J = 7.1, 9.7 Hz), 3.45 (d, 1 H, J = 14.1 Hz), 3.52 (d, 1 H, J = 14.1 Hz), 3.88 (t, 1 H, J = 6.4 Hz), 4.52 (dd, 1 H, J = 2.7, 9.7Hz); ¹³C-NMR (50 MHz, CDCl₃) δ 15.4 (q), 16.0 (q), 17.0 (q), 19.9 (q), 20.7 (q), 26.5 (t), 28.2 (d), 32.5 (d), 32.8 (t), 35.5 (t), 38.2 (t), 41.7 (d), 44.5 (d), 44.6 (s), 47.8 (s), 53.1 (t), 65.0 (d), 80.4 (d), 173.2 (s),
- 13) Compound 4 was identified by comparison (1 H-NMR, 13 C-NMR, IR, HRMS) with data of previous prepared samples. See for example: D. A. Evans, J. Bartroli, *Tetrahedron Lett.* **1982**, 23, 807. **4**: m.p. $122.5-123.5^{\circ}$ C, $[\alpha]_{D} = +41.3$ (c=2.1, CHCl₃).